

**CIHM
Microfiche
Series
(Monographs)**

**ICMH
Collection de
microfiches
(monographies)**



Canadian Institute for Historical Microreproductions / Institut canadien de microreproductions historiques

© 1998

Technical and Bibliographic Notes / Notes techniques et bibliographiques

The Institute has attempted to obtain the best original copy available for filming. Features of this copy which may be bibliographically unique, which may alter any of the images in the reproduction, or which may significantly change the usual method of filming are checked below.

- Coloured covers / Couverture de couleur
- Covers damaged / Couverture endommagée
- Covers restored and/or laminated / Couverture restaurée et/ou pelliculée
- Cover title missing / Le titre de couverture manque
- Coloured maps / Cartes géographiques en couleur
- Coloured ink (i.e. other than blue or black) / Encre de couleur (i.e. autre que bleue ou noire)
- Coloured plates and/or illustrations / Planches et/ou illustrations en couleur
- Bound with other material / Relié avec d'autres documents
- Only edition available / Seule édition disponible
- Tight binding may cause shadows or distortion along interior margin / La reliure serrée peut causer de l'ombre ou de la distorsion le long de la marge intérieure.
- Blank leaves added during restorations may appear within the text. Whenever possible, these have been omitted from filming / Il se peut que certaines pages blanches ajoutées lors d'une restauration apparaissent dans le texte, mais, lorsque cela était possible, ces pages n'ont pas été filmées.
- Additional comments / Commentaires supplémentaires:

Pagination is as follows: p. [570]-584.

L'Institut a microfilmé le meilleur exemplaire qu'il lui a été possible de se procurer. Les détails de cet exemplaire qui sont peut-être uniques du point de vue bibliographique, qui peuvent modifier une image reproduite, ou qui peuvent exiger une modification dans la méthode normale de filmage sont indiqués ci-dessous.

- Coloured pages / Pages de couleur
- Pages damaged / Pages endommagées
- Pages restored and/or laminated / Pages restaurées et/ou pelliculées
- Pages discoloured, stained or foxed / Pages décolorées, tachetées ou piquées
- Pages detached / Pages détachées
- Showthrough / Transparence
- Quality of print varies / Qualité inégale de l'impression
- Includes supplementary material / Comprend du matériel supplémentaire
- Pages wholly or partially obscured by errata slips, tissues, etc., have been refilmed to ensure the best possible image / Les pages totalement ou partiellement obscurcies par un feuillet d'errata, une pelure, etc., ont été filmées à nouveau de façon à obtenir la meilleure image possible.
- Opposing pages with varying colouration or discolourations are filmed twice to ensure the best possible image / Les pages s'opposant ayant des colorations variables ou des décolorations sont filmées deux fois afin d'obtenir la meilleure image possible.

This item is filmed at the reduction ratio checked below / Ce document est filmé au taux de réduction indiqué ci-dessous.

10x	14x	18x	22x	26x	30x
<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input checked="" type="checkbox"/>	<input type="checkbox"/>
12x	16x	20x	24x	28x	32x

The copy filmed here has been reproduced thanks to the generosity of:

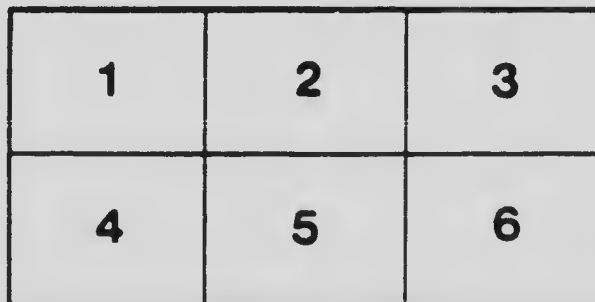
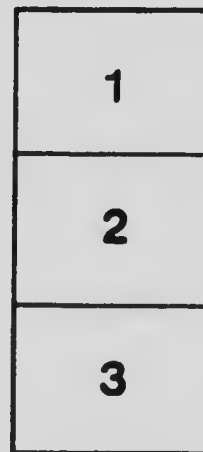
Engineering Sciences Library,
Queen's University

The images appearing here are the best quality possible considering the condition and legibility of the original copy and in keeping with the filming contract specifications.

Original copies in printed paper covers are filmed beginning with the front cover and ending on the last page with a printed or illustrated impression, or the back cover when appropriate. All other original copies are filmed beginning on the first page with a printed or illustrated impression, and ending on the last page with a printed or illustrated impression.

The last recorded frame on each microfiche shall contain the symbol \rightarrow (meaning "CONTINUED"), or the symbol ∇ (meaning "END"), whichever applies.

Maps, plates, charts, etc., may be filmed at different reduction ratios. Those too large to be entirely included in one exposure are filmed beginning in the upper left hand corner, left to right and top to bottom, as many frames as required. The following diagrams illustrate the method:



L'exemplaire filmé fut reproduit grâce à la générosité de:

Engineering Sciences Library,
Queen's University

Les images suivantes ont été reproduites avec le plus grand soin, compte tenu de la condition et de la netteté de l'exemplaire filmé, et en conformité avec les conditions du contrat de filmage.

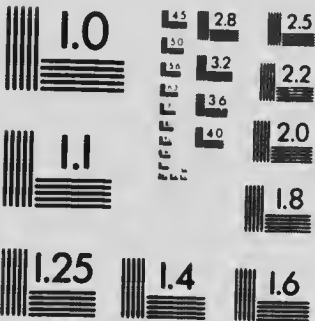
Les exemplaires originaux dont la couverture en papier est imprimée sont filmés en commençant par le premier plat et en terminant soit par la dernière page qui comporte une empreinte d'impression ou d'illustration, soit par le second plat, selon le cas. Tous les autres exemplaires originaux sont filmés en commençant par la première page qui comporte une empreinte d'impression ou d'illustration et en terminant par la dernière page qui comporte une telle empreinte.

Un des symboles suivants apparaîtra sur la dernière image de chaque microfiche, selon le cas: le symbole \rightarrow signifie "A SUIVRE", le symbole ∇ signifie "FIN".

Les cartes, planches, tableaux, etc., peuvent être filmés à des taux de réduction différents. Lorsque le document est trop grand pour être reproduit en un seul cliché, il est filmé à partir de l'angle supérieur gauche, de gauche à droite, et de haut en bas, en prenant le nombre d'images nécessaire. Les diagrammes suivants illustrent la méthode.

MICROCOPY RESOLUTION TEST CHART

(ANSI and ISO TEST CHART No. 2)



APPLIED IMAGE Inc

1651 East Main Street
Rochester, New York 14609 USA
(716) 482-0300 Phone
(716) 288-5989 Fax

UNIVERSITY OF TORONTO
STUDIES

PAPERS FROM THE CHEMICAL
LABORATORIES

No. 115: THE TOXICITY TOWARDS ANTHRAX AND
STAPHYLOCOCCUS OF SOLUTIONS CONTAINING PHENOL
AND SODIUM CHLORIDE, BY J. S. LEMON

(REPRINTED FROM THE JOURNAL OF PHYSICAL CHEMISTRY, VOL. XXIV)

THE UNIVERSITY LIBRARY: PUBLISHED BY
THE LIBRARIAN, 1920

University of Toronto Studies
COMMITTEE OF MANAGEMENT

Chairman: SIR ROBERT ALEXANDER FALCONER, LL.D., K.C.M.G.
President of the University

PROFESSOR W. J. ALEXANDER, PH.D.

PROFESSOR J. J. MACKENZIE, B.A., M.B.

PROFESSOR J. P. McMURRICH, PH.D.

BRIG.-GEN. C. H. MITCHELL, B.A.Sc., C.B., C.M.G., D.S.O.

PROFESSOR G. H. NEEDLER, PH.D.

PROFESSOR GEORGE M. WRONG, M.A.

General Editor: H. H. LANGTON, M.A.
Librarian of the University

THE TOXICITY TOWARDS ANTHRAX AND STAPHYLOCOCCUS, OF SOLUTIONS CONTAINING PHENOL AND SODIUM CHLORIDE

BY J. S. LEMON

The following experiments were carried out in the pathological laboratory of the University of Toronto in the winter of 1905-6, under the supervision of Prof. J. J. Mackenzie, with solutions supplied by Prof. W. Lash Miller.

Cultures of the "potato bacillus" (*B. mesentericus*) of anthrax (*B. anthracis*) and of *Staphylococcus pyogenes aureus* were grown on agar, the colonies washed off the agar, without breaking its surface, by 0.6 percent salt solution, and the suspension let stand (sometimes centrifuged) so that clumps might settle to the bottom. A second measured portion of the 0.6 percent salt solution was then infected by from one to three loops taken from the upper portion of the suspension; this constituted the "second suspension," in the case of potato spores and anthrax spores it was heated to 70° C to destroy vegetative forms. In the poisoning experiments, 10 cc of the toxic liquid was inoculated with a loopful of this "second suspension" and the time noted; to ensure that a good average sample should be removed for the inoculation, a mixing-rod was kept in the suspension tube, and the liquid was thoroughly stirred before a loopful was removed. The poison was then left to stand at room temperature, or in an incubator, and at measured intervals of time a loopful from it was added to 10 cc agar jelly, which had been melted and kept at 45° C; this was poured into a petri dish, allowed to solidify, and put away in the incubator. The number of colonies that grew on a measured area of the plate was taken as a measure of the number of cells left living in the poison at the time the agar was infected.

The usual precautions were taken against accidental infection—tubes and instruments were sterilized, tubes plugged with cotton wool which was "flamed" before removing, etc.,

and the loopfuls of suspension were measured out with the "machine."¹ "Neutral agar" holding 1.5 percent agar-agar was employed, and the cultures were in all cases grown on the same agar as that used for the plates, for fear there might be some difference in the preparations made up at different times—although in every case the same recipe was followed. Koch has shown the need for this precaution in his experiments with iodine, which he found would act well in a neutral medium containing a trace of albumin, but not nearly so actively in an alkaline medium containing more albumin.

Preliminary experiments were, of course, necessary in order that the number of colonies grown on a plate might be suitable for counting. In the case of staphylococcus, for instance, one cc of 0.6 percent brine was added to a 24-hour-old pure culture and shaken; the second suspension was made by adding three loops of the first to 5 cc of the brine, and 10 cc agar was infected with one loop of this second suspension. The plates were so thickly sown that a count could not be made. The procedure was varied by using 5 cc salt solution for the first suspension and 10 cc for the second, but the plates were still too thickly sown. Finally, by using 10 cc for the first and 10 cc for the second, a satisfactory count of 7300 colonies was obtained; the plates had an area of 63 to 64 square centimeters, and the number of colonies on 12 cm² of each plate was counted.

The spores were used instead of the vegetative forms of anthrax and potato bacillus because the former are much more resistant to phenol, and thus enabled the experiments to be made with higher concentrations of the poison. It was hoped that the innocuous potato spores might be used instead of the virulent anthrax, and preliminary experiments showed that insofar as their resistance is concerned, this is quite feasible; but it proved too difficult to prepare a uniform constant culture, while the very characteristic colonies of the anthrax made it easy to procure and maintain a pure strain, and in the end work with the potato bacillus was abandoned.

¹ Lash Miller: Jour. Phys. Chem., 24, 563 (1920).

Spores of the potato bacillus were obtained from a pure culture on agar, by incubating at 34°C over night and then leaving four days at room temperature ($19\text{--}20^{\circ}\text{C}$); anthrax spores by incubating the culture for 16 hours (when, according to Koeh spores begin to form) and then letting stand at room temperature for 48 hours. This procedure worked well in practice, though some observers place the optimum for spore production (anthrax) at 31°C and others at $24\text{--}35^{\circ}\text{C}$.

Resistance of the Spores to High Temperature

To compare their resistance to high temperatures, capillary tubes were filled with the second suspensions of potato spores, and anthrax spores respectively, their ends were sealed, and they were plunged into boiling water. After a measured interval they were removed, broken into sterile broth, and the growth noted. In the case of the potato spores there was a heavy growth after 2 minutes at 100°C , a smaller growth after 4 minutes, and a slight growth after 6 minutes, but none after 8, 10 or 15 minutes; in the case of anthrax, also, some were found alive after 6 minutes immersion in the boiling water.

Effect of Temperature on the Rate of Poisoning by Phenol

That the resistance of the potato spore to phenol is much the same as that of anthrax, may be seen by comparing the following measurements with those given later for anthrax spores; the immediate object of this set, however, was to ascertain how much effect the temperature of the poison bath had on the time it took the potato spores to die. Solutions were made up containing 2.5, 3.0, and 3.5 percent of phenol, respectively, and 10 cc of each of them was inoculated with one loop of the potato spore suspension, and kept at room temperature for two weeks, plates being poured each day. As a control, one loop of the same suspension was added to 10 cc of 0.6 percent salt solution at the same time as the others. Counts of plates made immediately after inoculation showed no less numbers than the control. On the second day, a very slight reduction in the number of colonies was observed on

As it seemed probable that the low counts might be due to the prolonged heating at 70° C, in the next and in all subsequent work the suspensions were heated to 70° C for 20 minutes only. The counts so obtained were much higher; but that may be due in part at least to a change in the way of counting, for in the first series colonies growing at a depth in the agar had not been counted, they differed in appearance from those on the surface and it was feared that the culture was not pure.

<i>B. anth.</i> in poison	25 ml.	1	2	3	4	5	6	7	hours
1.0% ph. + 5% NaCl (2.6)	153		75	130	55	57	77	104	49 colonies
3.0% phenol	182		174	101	88	97	89	75	56 colonies
3.5% phenol	195		125	40	87	83	36	36	colonies
2.5% ph. + 5% NaCl (3.6)	78		103	103	35	35	31	39	48 colonies
1.0% ph. + 10% NaCl (3.0)	137		118	60	21	45	25	11	21 colonies
4.0% phenol	103		107	118	44	30	50	35	25 colonies
2.5% ph. + 10% NaCl (5)	72		49	3	5	1	0	1	0 colonies

In the third series the suspension was thoroughly centrifuged after heating:

<i>B. anth.</i> in poison	20 ml.	1	2	3	4	5	6	7	8	hours
1.0% ph. + 5% NaCl (2.6)	142		134	114	84	46	18	13	23	0 colonies
3.0% phenol	124		81	109	73	34	15	18	17	9 colonies
3.5% phenol	119		107	90	48	12	17	10	5	0 colonies
2.5% ph. + 5% NaCl (3.6)	145		154	90	112	15	15	14	10	3 colonies
1.0% ph. + 10% NaCl (3.0)	115		92	41	51	7	5	7	2	0 colonies
4.0% phenol	124		110	110	53	19	18	13	15	2 colonies
2.5% ph. + 10% NaCl (5)	78		52	9	5	0	0	0	0	0 colonies

The fourth set were carried out under the same conditions as the third; the temperature of the incubator varied from 37.5° C to 42° C.

<i>B. anth.</i> in poison	15 m.	1	2	3	4	5	6	7	8	hours
1.9% ph. + 5% NaCl (2.6)	179	78	72	31	70	59	48	42	40	colonies
3.0% phenol	118	184	149	79	61	52	75	75	55	colonies
3.5% phenol	176	163	118	90	54	27	42	48	45	colonies
2.5% ph. + 5% NaCl (3.6)	179	78	72	31	70	59	48	42	40	colonies
1.9% ph. + 10% NaCl (3.9)	122	40	31	10	10	4	1	4	2	colonies
4.0% phenol	169	115	58	64	10	24	16	17	11	colonies
2.5% ph. + 10% NaCl (5)	90	16	0	0	0	0	0	0	0	colonies

Comparison of these four sets, shows that even when working with every care, duplicate results are not to be expected; the trouble lies, no doubt, in the uneven distribution of the spores in the suspensions from which measured volumes are taken for inoculation and for culture. The results, however, leave no doubt that the toxicities of the phenol-salt solutions are about equal to those of their salt-free equivalents.

A fifth set was undertaken, including 1.22 percent phenol with 10, 15 and 20 percent salt. In this case the culture after 16 hours in the incubator was kept for 6 days at room temperature, the growth washed off as usual with 10 cc 0.6 percent brine, heated in the tube of a centrifuge for 30 minutes to 70° C, agitated to break up clumps, and then centrifuged at high speed. Two loops of this (first) suspension were used to inoculate 10 cc of each of the poisons, and after standing in a water bath (whose temperature varied from 38° C to 44° C) for the time noted, one loop was used to infect the 10 cc of agar; the "machine" was used, and the same small platinum tube that was used in the experiments with staphylococcus.

With the exception of the solution containing 1.22 percent phenol and 15 percent salt, the order of toxicity is that of the equivalent solutions; the results obtained with this exceptional solution are abnormal also in the relatively large count after one hour, followed by a rapid decrease to zero.

<i>B. anth.</i> in poison	1(?)	2	3	4	5	6	7	8 hours
1.22% ph. + 10% NaCl (2.1)	1141	872	700	445	310	381	274	255 colonies
2.5% phenol	675	220	257	357	171	120	98	82 colonies
3.0% phenol	381	142	186	124	78	55	40	36 colonies
1.22% ph. + 15% NaCl (3.0)	117	40	10	2	3	0	0	0 colonies
3.5% phenol	140	106	41	25	25	16	13	11 colonies
4.0% phenol	58	14	34	32	16	8	8	— colonies
3.0% ph. + 5% NaCl	50	5	4	1	3	1	0	0 colonies
1.22% ph. + 20% NaCl (7+)	3	0	0	0	0	0	0	0 colonies

Effect of Salt on the Toxicity of Phenol towards *Staphylococcus*

This form was chosen for the experiments with lower concentrations of phenol. Colonies on agar, 24 hours old, were washed off with 0.6 percent brine, and suspensions made as already described; of course, the heating to 70° C was omitted. All infections were made with the "machine," using a small platinum tube as "loop." The poison acted at room temperature; 0.6 percent NaCl was used as control; the various cultures are distinguished by numbers.

Preliminary experiments with 1.0 percent phenol showed that the time required to kill all the cells was the same whether the poison was infected by a large or a small number of cells, and that a culture 47 hours old gave about the same results as one 24 hours old.

<i>Staph.</i> in poison	5	10	15	20	25	30	35	minutes
1.0% ph., cult. No. 4, 24.5° C	5	2	0	0	0	0	0	colonic
1.0% ph., cult. No. 5, 24.5° C	8	2	0	0	0	0	0	colonies
1.0% ph., cult. No. 6, 24.5° C	1402	29	0	0	0	0	0	colonies
1.0% ph., cult. No. 7, 24.5° C	512	5	0	0	0	0	0	colonies
1.0% ph., cult. No. 9, 24.5° C	2963	4	0	0	0	0	0	colonic
0.6% NaCl, cult. No. 7, 24.5° C	20491	18073	16514		15102			colonic
0.6% NaCl, cult. No. 9, 24.5° C	15082	15048	13048					colonies

With cultures Nos. 4 and 5, the machine was used; with the others the "biological lift," giving a much heavier infection. Culture No. 9 had been grown on agar for 47 hours; all the others in this paper for 24 hours only.

Experiments were then made to find the range of phenol concentrations within which poisoning takes place at a convenient rate; next the effect of sodium chloride, without phenol, was tested, and then comparison was made of the toxicity of 0.8, 0.7 and 0.6 percent phenol solutions with their chemical equivalents, viz., solutions containing 2.0 percent salt and 0.72, 0.63 and 0.54 percent phenol, respectively.

It soon became evident that experiments carried out under what were intended to be identical conditions, gave very different results. One cause of variation lay in the fact that equal volumes removed from the same suspension contain varying numbers of cocci—for instance, three plates prepared at the same time each from 10 cc of the same agar, infected in each case with the same volume of suspension (one loop taken with the machine), gave 5335, 5791, 6173 colonies, respectively, being a variation of 16 percent from the highest to the lowest. The principal cause, however, obviously lies in the variability of the staphylococcus itself, with culture No. 29 for instance, the time required for complete sterilization by 0.6 percent phenol was nearly twice as long as with culture No. 22, although in the first case the temperature was, if anything, a little higher.

In order to compare the toxicity of different solutions, therefore, it was necessary in every case to carry out simultaneous experiments with the same suspension; and as but little guidance could be obtained from previous experiments with the same poison but a different culture, a great many plates were poured which, on incubation, turned out to be sterile. All this added greatly to the amount of work and time required to obtain results; and if work of this kind is to be undertaken on any considerable scale, it will be necessary first to find some criterion of death which involves less delay than the plating method, and second to find conditions under which the

microbes experimented with can be grown "true to type" from the toxicological point of view. In comparison with the loss of time caused by the lack of these requisites, an uncertainty of 20 percent or so in the number of cells introduced into the poison is of little moment; and it is obvious that a more accurate regulation of temperature during the action of the poison can easily be attained by the use of a suitable thermostat.

SELECTION OF THE RANGE OF CONCENTRATIONS

Staph. No. 8 in poison									
	0	5	10	15	20	30	40	50	70 min.
0.25% phenol, 24.5° C	---	---	44977	---	---	55315	---	42995	59630 col.
0.50% phenol, 24.5° C	---	---	43984	---	51039	55032	49741	48129	---
1.0% phenol, 24.5° C	---	16927	73	0	---	---	---	---	---
0.6% NaCl, 24.5° C	39711	---	---	---	---	49724	---	---	43046 col.
Staph. No. 10 in poison									
	40	60	70	80	90	100	110	120 min.	
0.25% phenol, room temp.	---	---	7000	---	7573	---	6427	---	col.
0.5% phenol, room temp.	10500	7509	---	6637	---	6205	---	2418	col.
0.6% NaCl, room temp.	6045	---	---	---	---	7064	---	---	col.
Staph. No. 10 in poison									
	130	140	150	160	170 min.				
0.25% phenol, room temp.	5727	---	8591	---	7955	col.			
0.50% phenol, room temp.	---	1527	---	1209	---	col.			

In the experiments with culture No. 10, the room temperature varied from 20° C to 24° C, but as the tubes stood close together, and plates were poured from them alternately, the results are comparable.

Comparison of 0.80% Phenol with Its Chemical Equivalent

The equivalent contained 0.72 percent phenol and 2.0 percent salt. After "control" is given the number of colonies counted on plates from a 0.6 percent salt solution infected at the same time as the poison liquids; usually one plate from

this salt solution was poured immediately after inoculating, and another towards the close of the poison experiments.

Culture No. 17; temp. 22° C. Control: 7128, 8173 col.

Staph. in poison

5 6 10 11.5 15 16 20 21 30 40 50 67 min.

0.80% phenol 8210 -- 7191 -- 6084 -- 2864 734 44 0 0 col.

Equivalent

7058 -- 5728 -- 1819 -- 2149 -- 742 0 0 0 col.

Culture No. 18; temp. 20° C. Control: 8546 after 3 min., 10882 after 5 min.

Staph. in poison 5 10 15 20 30 40 55 60 70 min.

0.80% phenol 10426 8400 6428 3755 350 6 0 col.

Equivalent 10927 10692 6199 5543 2864 293 11 8 3 col.

Culture No. 29; temp. 24 27° C. Control: 10500 col.

Staph. in poison 5 10 15 20 30 40 55 70 min.

0.80% phenol 3309 309 1 0 0 0 0 col.

Equivalent 2546 55 38 4 0 0 0 col.

Towards culture No. 17, the phenol and its equivalent proved equally toxic; but towards Nos 18 and 29, the "equivalent" was less toxic than the pure phenol solution.

Comparison of 0.70% Phenol with Its Chemical Equivalent

The equivalent contained 0.63 percent phenol and 2.0 percent salt.

Culture No. 14; temp. 21° C. Control: 7828, 0555, 6493 col.

Staph. in poison

10.5 15 23 24.5 33 35 44 45 (11 more, to 98) min.

0.70% phenol 3384 -- 674 -- 142 3 0 col.

Equivalent

2262 -- 970 82 3 0 col.

The following experiment was carried out at the same time, with the same culture and controls, to compare the effect of 0.6 and 0.7 percent phenol.

Staph. in

poison 10 22.5 31 42 52.5 62.5 72.5 80 86.5 96.5 min.

0.6% ph. 5409 4975 3134 3039 2095 914 419 41 25 13 col.

Culture No. 14,
Staph. in

poison	14	15	23.5	24	34.5	35	44	45	etc., to 66 min.
0.70% phenol	—	1994	51	—	0	—	0	—	0 col.
Equivalent	1980	—	—	311	—	9	—	0	0 col.

Culture No. 16; temp. 22° C. Control: 5489 after 4 min., 5441 after 6 min.

Staph. in poison	10	20	30	40	50	60	70	85	100 min.
0.70% phenol	2896	980	22	2	0	0	0	0	0 col.
Equivalent	2604	2198	1604	1591	680	135	66	5	0 col.

Culture No. 27; temp. 23-24° C. Control: avg. 9673 colonies

Staph. in poison	11	21	30	40	50	60	75	90 min.
0.70% phenol	1273	11	0	0	0	0	0	0 col.
Equivalent	167	40	0	0	0	0	0	0 col.

Culture No. 30; temp. 19-25° C. Control: avg. 10163 colonies

Staph. in poison	10	20	30	40	50	60	75	90	105 min.
0.70% phenol	7937	1400	37	0	0	0	0	0	0 col.
Equivalent	1273	19	0	0	0	0	0	0	0 col.

The five experiments with culture No. 14 were carried out at the same time and under the same conditions; those with Nos. 27 and 30, together with the experiments on 0.60 percent phenol given below. Towards cultures 14 and 27 the phenol solution and its equivalent are equally toxic; towards No. 16 the equivalent was very much less toxic, while towards No. 30 the equivalent seems to be somewhat more toxic than the pure phenol solution, though the unsteadiness of the temperature renders this conclusion uncertain. In comparison with the difference between the death rates with 0.7 percent and 0.6 percent phenol, however, the 0.7 percent phenol and its chemical equivalent come very close together, except in the experiments with culture No. 16.

Comparison of 0.60% Phenol with Its Chemical Equivalent

The equivalent contained 0.54 percent phenol and 2.0 percent salt. Owing to the unexpected results obtained, a large number of experiments were made in the course of which fresh solutions were made up and the old ones were re-analyzed, the tubes, etc., used in the experiments were

exchanged for others, and every precaution taken to avoid accidental contamination; but the same general results were always obtained.

Culture No. 15; temp. 21° C. Control: 7471 after 2.5 min., 6627 after 5.5 min.

Staph. in poison

	10	20	30	40	50	60	70	82	97	117 min.
0.60% phenol										
Equivalent	5844	5791	5955	3914	2810	2535	1889	1623	616	57 col.
	7955	6351	7573	6151	5388	5476	5371	4484	1209	728 col.

Culture No. 19; temp. 23-27° C. Control: 3882, 5037 colonies

<i>Staph.</i> in poison	10	20	30	40	50	71.5	87.5	102.5	min.
0.60% phenol									
0.60% equivalent	576	0	0	—	0	0	0	0	0 col.
0.70% equivalent	3293	1591	1389	—	647	371	109	30	0 col.
	1718	1256	627	84	1	1	0	0	0 col.

Culture No. 20; temp. 23.5. Control: 5728, 6713 colonies.

Staph. in poison

	10	20	30	40	50	60	75.5	91	105	121 min.
0.60% phenol										
Equivalent	4935	858	6	2	0	0	0	0	0	0 col.
	6762	5473	3500	3946	3755	1009	1972	1095	389	170 col.

Culture No. 21; temp. 23.5° C. Control: 6619, 7636 colonies.

Staph. in poison

	10	20	30	40	50	60	75	90	105	120 min.
0.60% phenol (1)										
0.60% phenol (2)	1284	585	74	22	0	0	0	0	0	0 col.
Equivalent	763	—	1	0	0	0	0	0	0	0 col.
	7319	5728	4811	4694	4137	3710	1654	1600	1336	332 col.

Culture No. 22; temp. 22-24° C. Control: avg. 6470 colonies

Staph. in poison

	12	20	30	40	50	60	75	90	105	120 min.
0.60% phenol										
Equivalent	5728	2482	338	3	0	0	0	0	0	0 col.
	7433	6364	5855	5454	5405	5091	4614	2800	2609	1146 col.

Culture No. 23; temp. 23-27° C. Control: avg. 6250 colonies

Staph. in poison	10	20	30	40	50	60	75	90	105	120 min.
0.60% phenol (1)	4391	1745	1	0	0	0	0	0	0	0 col.
0.60% phenol (2)	3882	1852	317	2	0	0	0	0	0	0 col.
Equivalent	6619	5537	4968	4960	4028	3946	—	1728	1209	440 col.

Culture No. 24; temp. 22.5-25° C. Control: 9835, 10424 colonies

Staph. in poison	10	20	30	40	50	60	75	90	105	120 min.
0.60% phenol (1)	5727	4646	1146	104	2	0	0	0	0	0 col.
0.60% phenol (2)	7637	—	2819	—	40	1	0	0	0	0 col.
0.54% phenol	5307	4072	2545	500	3	0	0	0	0	0 col.
0.60% equivalent	8057	4455	—	1464	272	21	5	0	0	0 col.

Culture No. 25; temp. 20.5 24.5° C. Control: avg. 11137

Staph. in poison	10	20	30	40	50	60	75	90	105	120 min.
0.60% phenol	9164	7510	2927	305	1	0	0	0	0	0 col.
0.60% equivalent	8782	8591	4136	955	83	1	0	0	0	0 col.
0.54% phenol	8973	8591	7700	5918	580	109	5	0	0	0 col.

Culture No. 27; temp. 23 24° C. Control: avg. 9673 colonies

Staph. in poison	11	21	22	30	40	50	60	75	90	105 min.
0.60% phenol	9037	—	7828	6619	1856	1044	425	1	0	0 col.
Equivalent	7700	5155	—	2757	1686	636	283	63	29	0 col.

Culture No. 30; temp. 19 25° C. Control: avg. 10163 colonies

Staph. in poison	10	20	30	40	50	60	75	90	105 min.
0.60% phenol	10100	7541	6746	2227	573	127	4	0	0 col.
Equivalent	10100	5300	2227	11	39	15	1	0	0 col.

Towards Culture No. 30, where the unsteadiness of the temperature renders conclusions uncertain, the equivalent solution was more toxic than the 0.60 percent phenol. In every other case, the equivalent proved less toxic often very much less toxic than the pure phenol solution. In the case of No. 19, the solution containing 0.60 percent phenol was even more toxic than a solution containing more phenol and salt as well, viz., "the 0.70 percent equivalent" 0.61 percent

phenol + 2.0 percent salt; similarly with No. 24, a solution containing 0.54 percent phenol proved more toxic than one containing 0.54 percent phenol + 2.0 percent salt, but with No. 25, the reverse was the case. Towards culture No. 25, the 0.60 percent phenol was distinctly more toxic than the 0.54 percent phenol solution, while towards No. 24 there was no great difference, in the behaviour of the two solutions.

Summary

My experiments with anthrax spores show that the increased toxicity observed on adding salt to a phenol solution is in accordance with the assumption that two solutions of phenol, with or without salt, are equally toxic if their compositions are such that both would be in equilibrium with the same solution of phenol in toluene. The experiments with staphylococcus, however, in which lower concentration phenol were employed, show that while the assumption fairly in accord with the behaviour of 0.80 percent phenol, in the case of 0.60 percent phenol the chemically equivalent solution containing salt is much less toxic; 0.70 percent phenol occupies an intermediate position.

Every care has been taken to avoid accidental contamination of the vessels, and accidents in making up the solutions; and in view of the large number of corroborative experiments, the general result must be regarded as well established; but I have had no time to look farther into its cause.

The University of Toronto
June, 1920



UNIVERSITY OF TORONTO STUDIES

PAPERS FROM THE CHEMICAL LABORATORIES

No. 91: Experimental determination of binodal curves, plait points, and tie lines, in fifty systems, each consisting of water and two organic liquids, by WALTER D. BONNER.	0.25
No. 92: Mathematical theory of the changes of concentration at the electrode, Professors T. R. ROSEBRUGH and W. LASH MILLER	0.50
No. 93: The behaviour of copper anodes in chloride solutions, by S. DUSHMAN	0.25
No. 94: The chemical philosophy of the High School textbooks, by Professor W. LASH MILLER	0.25
No. 95: Lantern experiments on reactions in non-homogeneous systems, by FRANK B. KENRICK	0.25
No. 96: Some lantern experiments on surface tension, by FRANK B. KENRICK	0.25
No. 97: The phthalyl cyanides, by GIBBS BLACKSTOCK	0.25
No. 98: The influence of diffusion on electromotive force produced in solutions by centrifugal action, by Professor W. LASH MILLER	0.25
No. 99: Hyperbasis, by Professor FRANK B. KENRICK	0.25
No. 100: Electrodeposition of Metals, by Professor W. LASH MILLER	0.25
No. 101: Determination of free cyanide in cyanide copper and brass baths, by J. T. BURT-GERRANS and G. C. MORRISON	0.25
No. 102: The rate of dissociation of nitrogen peroxide, by W. L. ARGO	0.25
No. 103: Researches in Physical Chemistry, No. VII, by PROFESSORS W. LASH MILLER and FRANK B. KENRICK	0.25
No. 104: Friedel and Crafts' reaction—the preparation of orthobenzoyl-benzoic acid and benzophenone, by C. R. RUBIDGE and N. C. QUAA	0.25
No. 105: Studies on Filtration, by Professor J. W. BAIN and A. E. WIGLE	0.25
No. 106: The distribution of colloidal arsenious sulphide between the two liquid phases in the system water, ether, alcohol, by HARRY P. CORLISS	0.25
No. 107: On the formation of a badly conducting film on copper anodes in copper cyanide solutions, by W. LASH MILLER	0.25
No. 108: Orthobenzoyl-benzoyl chloride, by H. C. MARTIN	} 0.25
No. 109: The methyl ester of orthobenzoyl-benzoic acid, by T. C. McMULLEN	

No. 110: The action of a solution of potassium hydroxide in alcohol on oxalic esters, by N. C. QUA AND D. McLAREN.....	0.25
No. 111: The effect of chlorine on periodic precipitation, by Miss A. W. FOSTER	0.25
No. 112: The scattering of light by dust-free liquids, by W. H. MARTIN.....	0.25
No. 113: Friedel and Crafts' reaction—nitrophthalic anhydrides and acetylamino-phthalic anhydrides with benzene and aluminum chloride, by W. A. LAWRENCE.....	0.25
No. 114: Toxicity and chemical potential, by W. LASH MILLER	0.25
No. 115: The toxicity towards anthrax and staphylococcus of solutions containing phenol and sodium chloride, by J. S. L' MON.....	0.25

